Polysubstituted pyrans, chromenes, and chromenopyridines with isoxazole or isothiazole moiety: synthesis, structure, and antitumor activity

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SUPPLEMENTARY INFORMATION

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1. Methodology and results of biotesting

Materials and methods.

Media and components of culture media: medium DMEM 5648, antibiotics (penicillin, streptomycin, amphotericin B), fetal bovine serum (FBS), trypsin - EDTA (0,25% trypsin - 0,02% EDTA) (Sigma), dimethyl sulfoxide (DMSO).

Cell lines. Hela (cervical cancer, human), glioma *C6* (rat) from the collection of the Republican Research and Practical Center for Epidemiology and Microbiology, Belarus.

Study of the antitumor effect of compounds. Cells were seeded into wells of 96-well plates (Corning) in DMEM supplemented with 10% FBS and antibiotics (penicillin, streptomycin, amphotericin B). A day later, the test compounds were added to the wells. In the control, a solvent was added (dimethyl sulfoxide at a final concentration of 0,1%). Cultivated for 48 h at 37 °C and 5% CO₂. Then, cell samples were analyzed using the MTT assay with 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT - thiazolyl blue tetrazolium bromide, Glentham Life Sciences).

Medicines.

11a, **12e**, **15a**, **15c** – 0,5 mM stock solutions in 1% DMSO were prepared (first substances were diluted in DMSO, then brought to the required volume with isotonic sodium chloride solution). Further dilutions were made with 1% DMSO. Solutions were added to the wells of the plate with growing cells in a volume ratio 1 (**11a**, **12e**, **15a** μ **15c**) : 9 (medium with cells).

Carboplatin (Cp) is an alkylating drug, a platinum derivative. Forms cross-links between adjacent guanine pairs in DNA.

Doxorubicin (Dx) is an anthracycline antibiotic with antitumor action. Intercalates into DNA, disrupts DNA synthesis, and also causes other metabolic disorders in cells.

Cyclophosphamide (Ca) is an alkylating agent that alkylates DNA and proteins.

Fluorouracil (Fu) is an antimetabolite, an antagonist of pyrimidines (uracil). Disturbs DNA synthesis, suppresses RNA synthesis.

Ribomustine (Rm) (active component – bendamustine hydrochloride) is an alkylating drug that disrupts the structure and synthesis of DNA.

Results.

Substances **11a**, **12e**, **15a** and **15c** at a concentration of 200 μ M suppressed the growth of *C6* glioma cells (Fig. S1), substances **11a**, **12e** – the growth of *Hela* cells (Fig. S2). Carboplatin at a concentration of 10–40 μ M had a dose-dependent inhibitory effect on the growth of tumor

cells. The combined use of compound **11a** with carboplatin led to an enhancing of inhibitory effect. A similar but less pronounced effect was observed when carboplatin was combined with the compound **12e**; the same trend was observed when **12e** acted on *Hela* cells. Co-administration of carboplatin with compound **15a** provided some additional effect only at carboplatin concentrations of 20 and 40 μ M, Fig. S2. Co-administration of carboplatin with compound **15c** resulted in a weak tendency to increase the effect compared to compound **15c**.



Fig. S1. Effect of carboplatin at a concentrations of 10–40 μ M and substances **11a**, **12e** (a) and **15a**, **15c** (b) at a concentration of 200 μ M on the growth of *C6* glioma cells; *p < 0,05 when compared with control; **p < 0,05 when compared with the effect of carboplatin; ***p < 0,05 when compared with the effect of a heterocyclic compound (Mann-Whitney test)



Fig. S2. Effect of carboplatin at a concentrations of 10–40 μ M and substances **11a**, **12e** (a) and **15a**, **15c** (b) at a concentration of 200 μ M on the growth of *Hela* cells; *p < 0,05 when compared with control; **p < 0,05 when compared with the effect of carboplatin; ***p < 0,05 when compared with the effect of a heterocyclic compound (Mann-Whitney test)

Carboplatin at low doses (5 and 0,5 μ M) didn't affect the growth of *C6* glioma and *Hela* cells. At low doses of carboplatin (5 and 0,5 μ M) substances **11a**, **12e**, **15a** and **15c** showed their own inhibitory effect against *C6* glioma cells, but no additional effect was detected. There was a weak tendency to enhance the effect of 5 μ M carboplatin when combined with **15a** on *C6* glioma cells (Fig. S3a) and a tendency to enhance the effect of 5 μ M carboplatin when combined with **12e**, **15a**, **15c** on *Hela* cells (Fig. S3b). In this and the next experiment, compounds **11a**, **12e**, **15a** and **15c** inhibited the growth of *C6* glioma cells more significantly than in the previous experiment. This could be because of some variation in the onset of the exponential phase of cell growth from experiment to experiment. It can be assumed that conducting the experiment in the most rapid growth phase contributes to a better expression of the effect of growth-inhibiting drugs.



Fig. S3. Effect of carboplatin at a concentrations of 5 and 0,5 μ M and substances **11a**, **12e**, **15a** and **15c** at a concentration of 200 μ M on the growth of *C6* glioma (a) and *Hela* (b) cells; *p < 0,05 when compared with control; **p < 0,05 when compared with the effect of a heterocyclic compound; ***p < 0,05 when compared with the effect of carboplatin (Mann-Whitney test)

Compounds **11a**, **12e**, **15a** and **15c** at a dose of 100 μ M with carboplatin at a concentration of 40 μ M tended to show a weak additional effect against *C6* glioma cells (Fig. S4a). For *Hela* cells, a similar effect was observed when using **11a** or **12e** (Fig. S4b).



Fig. S4. Effect of carboplatin at a concentration of 40 μ M and substances **11a**, **12e**, **15a** and **15c** at a concentration of 100 μ M on the growth of C6 glioma (a) and *Hela* (b) cells; *p < 0,05 when compared with control; **p < 0,05 when compared with the effect of a heterocyclic compound; ***p < 0,05 when compared with the effect of carboplatin (Mann-Whitney test)

An experiment with ribomustine was conducted taking it in a small dose of 5 and 0,5 μ M. At such doses it had almost no effect on cell growth. The combined use of 5 μ M ribomustine with **11a**, **12e**, **15a** and **15c** didn't lead to a significant change in the effect compared to the effect of heterocyclic compounds, and when using 0,5 μ M ribomustine together with **12e**, **15a** and **15c**, a tendency to enhance the effect was observed (Fig. S5). We have already noted a similar phenomenon of increasing synergistic effect with decreasing dose.¹



Fig. S5. Effect of ribomustine at a concentrations of 5 and 0,5 μ M and substances **11a**, **12e**, **15a** and **15c** at a concentration of 200 μ M on the growth of *C6* glioma (a) and *Hela* (b) cells; *p < 0,05 when compared with control; **p < 0,05 when compared with the effect of a heterocyclic compound; ***p < 0,05 when compared with the effect of ribomustine (Mann-Whitney test)

An experiment was conducted on *C6* glioma cells using other chemotherapy drugs: doxorubicin, cyclophosphamide and fluorouracil. Compound **11a** had a strong inhibitory effect;

S5

against this background, no enhancement of the effect of chemotherapy drugs was detected (compared to **11a**). Derivatives **12e** and **15a** enhanced the effect of doxorubicin, cyclophosphamide and fluorouracil (Fig. S6a, b), and **15c** caused a weak tendency to increase the effect (with doxorubicin and uracil) or had no effect (with cyclophosphamide) (Fig. S6b).



Fig. S6. The effect of combined use of doxorubicin, cyclophosphamide and fluorouracil and compounds **11a**, **12e** (a) and **15a**, **15c** (b) at a concentration of 200 μ M on the growth of *C6* glioma cells; *p < 0,05 when compared with control; ** p < 0,05 when compared with the effect of a chemotherapy drug; *** p < 0,05 when compared with the effect of a heterocyclic compound (Mann-Whitney test)

Conclusion

1. Compounds **11a**, **12e**, **15a**, **15c** can have an inhibitory effect on the growth of tumor cells. The most active one is **11a**, the least active is **15c**.

2. The combined use of compounds **11a**, **12e**, **15a** with chemotherapy drugs leads to an increased inhibitory effect on tumor cells compared to the action of the drugs alone. Compound **15c** weakly enhances or doesn't enhance the effects of drugs.

3. The effects of 11a, 12e, 15a, 15c are similar for C6 glioma and Hela cells.

4. Experiments with ribomustine revealed an effect of increasing the synergistic effect of compounds **12e**, **15a**, **15c** with a decrease in the dose of the chemotherapy drug.

2. X-Ray diffraction analysis

X-ray diffraction experiments were carried out on an automatic four-circle area-detector diffractometer Bruker KAPPA APEX II (MoK α radiation).² The unit cell constants were refined over the whole data set.³ The experimental intensities were corrected for absorption using the SADABS program.⁴ The structure was solved by the intrinsic phasing method (SHELXT⁵) and refined by the full-matrix least squares method (SHELXL-2018/3⁶) on F^2 for all data in the anisotropic approximation for all non-hydrogen atoms. The H atoms of CH, CH₂, and CH₃ groups were introduced at geometrically calculated positions with $U_{iso}(H) = 1.2U_{eq}(C)$ for CH, and CH₂ groups and $U_{iso}(H) = 1.5U_{eq}(C)$ for CH₃ ones. The H atoms of NH₂ group in **9b** were introduced at geometrically calculated positions with $U_{iso}(H) = 1.2U_{eq}(N)$, in **11a** and **12d** the H atoms of NH₂ group were refined with $U_{iso}(H) = 1.2U_{eq}(N)$. The structure **11a** contains, contains, presumably, a half of the disordered dichloromethane molecule per formula unit. The PLATON program package.⁷

The main crystallographic data and characteristics of X-ray diffraction experiment are given in Table S1. The atomic coordinates have been deposited with the Cambridge Crystallographic Data Centre, depositions CCDC 2333536–2333538.

Identification code	9b	11a	12d
CCDC deposition number	2333536	2333537	2333538
Empirical formula	$C_{17}H_{18}Cl_2N_2O_4S$	$C_{13.5}H_{10}Cl_3N_3O_2S$	$C_{21}H_{19}N_3O_3$
Formula weight	417.29	384.65	361.39
Temperature/K	100(2)	100(2)	100(2)
Crystal system	monoclinic	monoclinic	monoclinic
Space group	C2/c	P2/c	C2/c
a/Å	27.927(3)	11.7288(10)	26.3449(18)
b/Å	8.1910(10)	8.8967(8)	8.6497(6)
c/Å	20.626(4)	16.6618(14)	16.8120(11)
α/°	90	90	90
β/°	129.048(3)	107.973(4)	104.961(3)
γ/°	90	90	90
Volume/Å ³	3664.3(9)	1653.8(3)	3701.2(4)
Z	8	4	8
$\rho_{calc}g/cm^3$	1.513	1.545	1.297
µ/mm ⁻¹	0.494	0.690	0.089

Table S1. Crystal data and structure refinement.

F(000)	1728.0	780.0	1520.0
Crystal size/mm ³	$0.5 \times 0.06 \times 0.03$	$0.36 \times 0.2 \times 0.06$	$0.18 \times 0.16 \times 0.08$
Radiation	MoKα (λ = 0.71073)	MoKα (λ = 0.71073)	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/°	7.516 to 59.998	6.886 to 59.998	8.674 to 59.994
Index ranges	$-39 \le h \le 38, -9 \le k \le 11,$	$-16 \le h \le 16, -12 \le k \le 12,$	$-36 \le h \le 36, -12 \le k \le 12,$
index ranges	$-28 \le l \le 28$	$-20 \le l \le 23$	$-23 \le l \le 23$
Reflections collected	40315	28436	35141
	5334	4824	5379
Independent reflections	$[R_{int} = 0.1694, R_{sigma} =$	$[R_{int} = 0.0892, R_{sigma} =$	$[R_{int} = 0.0828, R_{sigma} =$
	0.1371]	0.0749]	0.0720]
Data/restraints/parameters	5334/0/235	4824/0/196	5379/0/252
Goodness-of-fit on F ²	1.001	1.021	1.025
Final R indexes [I>=2 σ	$R_1 = 0.0564, wR_2 =$	$R_1 = 0.0464, wR_2 =$	$R_1 = 0.0619, wR_2 =$
(I)]	0.0988	0.0981	0.1446
Final P indexes [all data]	$R_1 = 0.1469, wR_2 =$	$R_1 = 0.0845, wR_2 =$	$R_1 = 0.1230, wR_2 =$
	0.1260	0.1118	0.1767
Largest diff. peak/hole / e Å ⁻³	0.41/-0.47	0.43/-0.36	0.58/-0.27



Figure S7. H-Bonding in 9b.

Table S2. Hydrogen bonds for 9b.

D	Н	Α	d(D-H)/Å	d(H-A)/Å	d(D-A)/Å	D-H-A/°
N1	H1A	03	0.88	2.07	2.691(3)	127
N1	H1B	N12 [*]	0.88	2.15	3.008(4)	164

CI CI CI N O S

*1-X,1-Y,1-Z



Figure S8. H-Bonding in 11a.

Table S3. Hydrogen bonds for 11a.

D	Н	Α	d(D-H)/Å	d(H-A)/Å	d(D-A)/Å	D-H-A/°
N1	H1A	N2*	0.82(2)	2.18(2)	2.991(3)	172(2)
N1	H1B	O2 ^{**}	0.87(2)	2.06(2)	2.911(2)	167(2)

*-X,2-Y,1-Z; **+X,1-Y,1/2+Z





Figure S9. H-Bonding in 12d.

 Table S4. Hydrogen bonds for 11d.

D	Н	Α	d(D-H)/Å	d(H-A)/Å	d(D-A)/Å	D-H-A/°
N2	H2A	N3 [*]	0.85(2)	2.21(3)	3.054(3)	169(2)
N2	H2B	O5 ^{**}	0.91(3)	1.99(3)	2.868(2)	162(2)

*1-X,-Y,-Z; **+X,1-Y,-1/2+Z

3. References

- 1. Potkin V.; Pushkarchuk A.; Zamaro A.; Zhou H.; Kilin S.; Petkevich S.; Kolesnik I.; Michels
- D.; Lyakhov D.; Kulchitsky V. SciRep. 2023, 13, 13624.
- 2. Apex2 // Bruker AXS Inc., Madison, Wisconsin, USA (2008).
- 3. SAINT-Plus (Version 8.40B) // Bruker AXS Inc., Madison, Wisconsin, USA (2019).
- 4. Krause L.; Herbst-Irmer R.; Sheldrick G.M.; Stalke D. J. Appl. Cryst. 2015, 48, 3.
- 5. Sheldrick G.M. Acta Crystallogr. 2015, A71, 3.
- 6. Sheldrick G.M. Acta Crystallogr. 2015, C71, 3.
- 7. Spek A.L. Acta Crystallogr. 2015, C71, 9.

4. Copy of the NMR spectra and the data of mass-spectrometry





Data Filename Sample Type Instrument Name	PS-28 Samp Instru	363_01.d le ument 1	Sample Name Position User Name	e PS-2863 Vial 2
Acq Method IRM Calibration Status Comment	All_20	D21_kol 1-6.m Applicable	Acquired Tim DA Method	e 7/1/2021 10:42:48 AM ChromPeakSurvey-Default.m
Sample Group Stream Name	LC 1	Info. Acquis Versio	sition SW	6400 Series Triple Quadrupole 10.0 (127)

User Chromatograms

CI

2a

CI



1.





m/z	Z	Abund
133.1	1	84212.9
277	1	123568.3
278	1	16629.91
279	1	120893.27
280	1	15571.5
281	1	38766.65
299	1	72328.09
301	1	100666.72
302	1	14271.13
303	1	22930.73

Spectrum Source

Peak (1) in "DAD1 - B:Sig=220,4 Ref=off"



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1.



192.265 137.447 133.910 130.191 129.477 126.216 126.079 121.056 143.741 46.240 20.248 19.940 **2b** CI CI CI 11 . hy many show with the off and metal for the hour and many and the second and the second second of the second of th where the managed and a ship of the second statement of the atractional and a second a 200 150 100 50 ppm (t1)

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Data Filename Sample Type Instrument Name Acq Method IRM Calibration Stat Comment	tus	EPIO_01.d Sample Instrument 1 All_2021_kol 1-6.m Not Applicable	Sample N Position User Nan Acquired DA Methe	lame ne Time od	EPIO Vial 3 3/21/2024 12:25:20 P QualDAMethod.m	М
Sample Group Stream Name	LC 1		Info. Acquisition SW Version	64(Ou	00 Series Triple adrupole 10.0 (127)	

Version

User Chromatograms

_<mark>O</mark>H =N

3a





Ionization Mode Collision Energy Fragmentor Voltage Spectrum Source 0 ESI 100 Peak (1) in "+ TIC Scan"

OH =N 3a \square

Qualitative Analysis Report





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τ. 139.957 139.709 137.892 124.669 123.793 170.276 130.763 127.192 159.681 19.935 19.843 96.963 3b ЮН .` 50 150 100 ppm (t1)

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:D:\msdchem\1\DATA\PV2_0721\ZV_949.D
File
Operator
           :
          : 20 Jul 2022 9:47 pm using AcqMethod BA 1SL.M
Acquired
Instrument :
               Instrument #1
Sample Name: ZV-949
Misc Info :
Vial Number: 8
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ppm (t1)

Quadrupole 10.0 (127)

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IRM Calibration St	atus	Not Applicable	DA Method	ChromPeakSurvey-Default.m
Comment				
Sample Group		1	Info.	
Stream Name	LC 1		Acquisition SW 6	400 Series Triple

Version

User Chromatograms

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4c



Integration Peak List









Spectrum Source

Peak (1) in "DAD1 - B:Sig=220,4 Ref=off"



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Operator
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Acquired
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Instrument :
               Instrument #1
Sample Name: ZH-377
Misc Info :
Vial Number: 1
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File :C:\msdchem\1\DATA\PV8_0313\PS_2543.D
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Instrument : Instrument #1
Sample Name: <u>PS-2543</u>
Misc Info :
Vial Number: 1
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Instrument : Instrument #1
Sample Name: <u>PS-2546</u>
Misc Info :
Vial Number: 3
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h

^{*}¹H NMR (500 MHz, CDCl₃) δ = 7.75 (d, J=8.3, 1H), 7.32 (d, J=8.4, 1H), 4.42 (q, J=7.1, 1H), 2.71 (q, J=7.6, 1H), 1.41 (t, J=7.1, 2H), 1.27 (t, J=7.6, 2H).



5C







-128.84







¹H NMR (500 MHz, CDCl₃) δ = 7.25 (d, J=7.8, 1H), 4.42 (q, J=7.1, 2H), 1.42 (t, J=7.1, 3H).

.

5d CO₂Et



100 BE 10 100 100 100 100

SK_zh3_03222023

172.73

bbo_13CF_bar CDCl3 /v nmrsu 15



5d CO₂Et CN









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Sample Type	Unavailable	Position	Unavailable	
Instrument Name	Unavailable	User Name	Unavailable	
Acq Method		IRM Calibration Status	Success	
DA Method	Default.m	Comment	Sample information is unavailable	[M] = 221

User Chromatograms



Integration Peak List

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v10.3	DAD1 - A	A:Sig=21	0,4 PS25	32 01.d							1
1 5		1		_		1.1.	* 6 667				
1.5							0.007				
1.25											
1-											
0.75											
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0.25											
0-			_								
) 0.5	1 1.5	2 2.5	5 3 3.5 Respons	4 4.5 5 e Units vs. Ad	5.5 6 cquisition Tim	6.5 7 ie (min)	7.5 8	3 8.5	99.	5

Integration Peak List

Peak 3	Start	RT	End	Height	Area	Area %
1	6,513	6,667	6,807	1734,33	7705,43	100

User Spectra





m/z	z	Abund.
105		84578
140		54485
146		103453
167		73977
174	1	222866
175	1	55138
194		234144
222	1	842317
223	1	118929
240		153418

Spectrum Source

Peak (1) in "DAD1 - A:Sig=210,4"



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Data Filename	TS9_k1.d	Sample Na
Sample Type	Unavailable	Position
Instrument Name	Unavailable	User Name
Acq Method		IRM Calibra
DA Method	Default.m	Comment

Sample Name Position User Name IRM Calibration Status Comment



Sample information is unavailable

User Chromatograms



Integration Peak List



Integration Peak List

Peak	Start	RT	End	Height	Area	Area %
1	5,547	5,673	5,780	416,69	1863,74	100

User Spectra





m/z	z	Abund.
236,1	1	1143691
237,1	1	172393
254,1	1	532860
255,1	1	96979

Spectrum Source



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D:\Ira\Spectra...0\ZH8.D\DATA.MS Injection 1 Function 1 (ZH8) TIC

110



¹¹¹H NMR (500 MHz, CDCl₃) δ = 7.55 (d, *J*=7.9, 1H), 7.26 (d, *J*=7.8, 2H).



SK_zh-7_03292023







127.286 126.185 119.953 113.279 131.048 129.760 160.140 157.299 198.045 170.111 167.367 31.902 30.492 19.424 54.341 99.443 7a 0 CN Me NH₂ Me 0 2 contraction of the association of the second 50 100 150 200

Data Filename PS2813_01.D **Sample Type** Sample **Instrument Name** Acq Method **IRM Calibration Status** Comment

Instrument 1 ALL_2020_KOL2.M Not Applicable

True

Sample Name Position **User Name Acquired Time DA Method**

Vial 2

10/15/2020 2:16:09 PM ChromPeakSurvey-Default.m

7a Me Me NH₂

OperatorName RunCompletedFlag

User Chromatograms

Fragmentor Voltage 135 **Collision Energy** 0 **Ionization Mode** ESI +ESI TIC Scan Frag=135.0V PS2813_01.D x10 6 * 7.574 1.75 1.5 1.25 1 0.75 0.5 0.25 0 5 4 4.5 5 5.5 6 6.5 7 Counts vs. Acquisition Time (min) 7.5 0.5 1.5 2 2.5 8 8.5 9 1 3 3.5 7 9.5 10

Integration Peak List

Peak	Start	RT	End	Height	Area	Area %
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Integration Peak List

Peak	Start	RT	End	Height	Area	Area %
1	7,246	7,493	7,813	2526,35	20151,26	100

User Spectra

Spectrum Source	Fragmentor Voltage	Collision Energy	Ionization Mode
Peak (1) in "+ TIC Scan"	135	0	ESI



Spectrum Source

1

1

665.3

666.3

Peak (1) in "DAD1 - A:Sig=210,4 Ref=off"

45190.74 18744.09



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Data Filename		ps-2921_001.d	Sample Name	ps-2921	7b 🦳
Sample Type		Sample	Position	Viai 2	(/
Instrument Name Acq Method IRM Calibration St Comment	tatus	Instrument 1 All_2021_kol 1-2.m Not Applicable	User Name Acquired Time DA Method	e 5/11/2022 12:16:18 PM Defaul1t.m	
Sample Group Stream Name	LC 1	In Ac Vé	fo. equisition SW	6400 Series Triple Quadrupole 10.0 (127)	Me O Nn ₂

User Chromatograms



0.5 1 1.5 2 2.5 3 3.5 4 4.5 5 5.5 6 6.5 7 7.5 8 8.5 Response Units vs. Acquisition Time (min)

Integr	ation	tart	RT	End	Height	Area	Area %	
reak	1	4,308	4,448	4,815	1946,31	13879,29	100	
User	Spe	ectra						Terization Mode

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User Chromatograms







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Data Filename		I-737_01.d	Sample Nam	ie I-737	7d /
Sample Type		Sample	Position	Vial 2	10
Instrument Name		Instrument 1	User Name		
Acq Method		All_2021_kol 1-2.m	Acquired Tir	me 5/29/2023 11:15:48 AM	○ ──
IRM Calibration St	atus	Not Applicable	DA Method	Defaul1t.m	0 ^N *
Comment					Me
Sample Group			Info.		Me NH ₂
Stream Name	LC 1		Acquisition SW Version	6400 Series Triple Quadrupole 10.0 (127)	

User Chromatograms



0.4

0.2 0

0.5	1	15	2	25	3	3.5	4	4.5	5	5.5	6	6.5	7	7.5	8	8.5	9
0.0		1.0	_		Res	ponse	Units	s vs. A	cquis	sition 1	Time	(min)					

* 6.679

1 6,579 6,679 6,905 306,04 2022,04 100 User Spectra	Peak	Start	RT	End	Height	Area	Area %	
User Spectra	1	6,579	6,679	6,905	306,04	2022,04	100	
	User S	pectra						

Spectrum Source	Fragmentor voltage	Collision chergy	aviiiad civii i iouo
Peak (1) in "+ TIC Scan Sub"	135	0	ESI

٢.,



---- End Of Report ----

SK_i709_12212022

bbo_1H_bar CDCl3 /v nmrsu 14

¹H NMR (500 MHz, CDCl₃) δ = 5.04 (d, *J*=0.8, 1H), 4.05 (m, 1H), 2.35 (m, 1H), 1.12 (t, *J*=7.1, 2H).



12



SK_i709_12212022









--- End Of Report ---

٢.,

SK_i705_12142022

bbo_1H_bar CDCl3 /v nmrsu 3

¹H NMR (500 MHz, CDCl₃) δ = 4.06 (q, *J*=7.0, 1H), 2.21 (dd, *J*=39.9, 16.2, 1H), 1.14 (t, *J*=7.1, 1H).



C1.10







User Chromatograms



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--- End Of Report ---

¹H NMR (500 MHz, DMSO) δ = 4.36 (t, *J*=4.8, 1H), 2.61 (t, *J*=5.9, 2H).



SK_i723_03222023

bbo_13CF_bar DMSO /v nmrsu 7



Data Filename	I 723_01.d	Sample Nar	me I 723	11a 🛛
Sample Type	Sample	Position	Vial 3	s i i ca
Instrument Name	Instrument 1	User Name		N CI
Acq Method	All_2021_kol 1-2.m	Acquired Ti	me 3/30/2023 12:37:20 PM	
IRM Calibration Status	Not Applicable	DA Method	Defaul1t.m	
Comment				O NH
Sample Group		Info.		
Stream Name LC 1		Acquisition SW	6400 Series Triple	
		Version	Quadrupole 10.0 (127)	

User Chromatograms



Integration Peak List



User Spectra

Spectrum Source	Fragmentor Voltage	Collision Energy	Ionization Mode
Peak (1) in "+ TIC Scan"	135	0	ESI



Peak List m/z z Abund 189.1 544201.94 1 3760455.75 364 1 [M+Na]+ 365 498165.03 1 366 1 2555208 367 1 341198.34 368 1 426182.44 705 844164.31 707 1249904.13 708.1 362644.34 709 1 674271.63



--- End Of Report ---

SK_i722_03222023

bbo_1H_bar DMSO /v nmrsu 5

¹H NMR (500 MHz, DMSO) $\delta = 2.59$ (dd, *J*=17.6, 1.2, 1H), 2.42 (d, *J*=17.6, 1H), 2.30 (d, *J*=16.1, 1H), 2.09 (d, *J*=15.8, 1H).



1.05



SK_i722_03222023

bbo_13CF_bar DMSO /v nmrsu 5





User Chromatograms



User Spectra

And it is not an		And the canonical statics of storage and second static station	
Spectrum Source	Fragmentor Voltage	Collision Energy	Ionization Mode
Peak (1) in "+ TIC Scan"	135	0	ESI

CN

NH2





1.5 0.5

280 300 320 mAU vs. Wavelength (nm)





Data Filename Sample Type **Instrument Name** Acq Method **DA Method** Default.m

PS2539R_k1.d Unavailable Unavailable

Sample Name Position **User Name IRM Calibration Status** Comment

Unavailable Unavailable Unavailable Success Sample information is unavailable

12a

CN

NH₂

User Chromatograms



Integration Peak List



Integration Peak List

Peak	Start	RT	End	Height	Area	Area %
1	1,213	1,340	1,507	161,41	1022,17	100

User Spectra





--- End Of Report ---





Data File Sample Instrum Acq Met DA Meth	ename Type Ient Name thod hod	PS25 Unav Unav Defa	35_01.d ailable ailable ult.m	Samı Posit User IRM Com	ile Name ion Name Calibration S ment	tatus	Unavailable Unavailable Unavailable Success Sample inf unavailable	e e formation e	n is		12b
User C	hromato	grams					Eci				
Frag	gmentor Vol	tage	135	Collision Energy	0 101	hization Mode					
x10 ⁷ 3.5-	+ IIC Sci 1	an PS25	35_01.a	* 4.792							1
3- 2.5- 2											
1 5											
1.5-											
0.5				- mark				_			
ntegratio Peak	ion Peak Lis	t RT 4 792	End 4 937	Height	Area 159197249	Area %					
	4,710	ч,/ 92									
			0 4 PS2	535 UI.a							
x10 ³	DAD1 - E	3:Sig=22	0,1102	* 4 740							
x10 ³	DAD1-E	3:Sig=22	0,1102	* 4.740							
x10 ³ 1.2	DAD1 - E	3:Sig=22	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	* 4.740							
x10 ³ 1.2 1 0.8		3:Sig=22		* 4.740							
x10 ³ 1.2 1 0.8 0.6		3:Sig=22	.,	* 4.740							
x10 ³ 1.2 1 0.8 0.6 0.4		3:51g=22		* 4.740							
x10 ³ 1.2 1 0.8 0.6 0.4 0.2		3:Sig=22		* 4.740							
x10 ³ 1.2 1 0.8 0.6 0.4 0.2 0		3:Sig=22		* 4.740							
x10 ³ 1.2 1 0.8 0.6 0.4 0.2 0) 1	2	3	* 4.740 * 4.740	6 e Units vs. A	7 8 .cquisition Tiu	9 me (min)	10	11	12	13
x10 ³ 1.2 1 0.8 0.6 0.4 0.2 0 Integrati) 1	2:51g=22	3	* 4.740 * 4.740 4 5 Respons	6 e Units vs. A	7 8 .cquisition Tir	9 me (min)	10	11	12	13
x10 ³ 1.2 1 0.8 0.6 0.4 0.2 0 Integrati Peak) 1 Start	2 st RT	3 End	* 4.740 * 4.740 4 5 Respons Height 7 1430 75	6 e Units vs. A Area 4297 97	7 8 cquisition Tir	9 me (min)	10	11	12	13



Agilent Technologies

Printed at: 14:52 on:16.03.2018

m/z	z	Abund.
189,1	1	796983
190,1	1	84013
348,2	1	967384
349,2	1	206095
540,8		106456
714,4		65592
717,2		175214
718,3		80006

Spectrum Source

Peak (1) in "DAD1 - B:Sig=220,4"



--- End Of Report ---

12b

CN



bbo_1H_bar DMSO /v nmrsu 11

¹H NMR (500 MHz, DMSO) δ = 7.74 (d, J=8.1, 1H), 7.34 (d, J=8.1, 1H), 1.19 (t, J=7.6, 1H).

C (d)	B (d



12c

O N

0

CN

NH₂

A (t) 1.19



Data Filename Sample Type		ZH-12_t1.d Sample	Sample Name Position	ZH-12 Vial 2	12c
Instrument Name Acq Method IRM Calibration St	atus	Instrument 1 All_2021_kol 1-2.m Not Applicable	User Name Acquired Time DA Method	5/12/2023 10:45:01 AM Defaul1t.m	o N
Comment		Taf	0		CN
Sample Group Stream Name	LC 1	Acc	quisition SW 64	400 Series Triple uadrupole 10.0 (127)	O NH ₂

User Chromatograms





Spectrum Source





--- End Of Report ---

SK_zh11_04192023

bbo_1H_bar DMSO /v nmrsu 2

¹H NMR (500 MHz, DMSO) δ = 7.54 (d, *J*=7.8, 1H), 7.26 (d, *J*=7.9, 1H), 2.62 (t, *J*=5.8, 2H).





Data Filename Sample Type		ZH-11_t1.d Sample	Sample Nam Position	e ZH-11 Vial 2	12d
Instrument Name Acq Method IRM Calibration Sta Comment	tus	All_2021_kol 1-2.m Not Applicable	Acquired Tin DA Method	ne 5/12/2023 10:22:29 AM Defaul1t.m	
Sample Group Stream Name	LC 1	I A V	nfo. cquisition SW ersion	6400 Series Triple Quadrupole 10.0 (127)	O NH ₂

User Chromatograms





--- End Of Report ---




Data File Sample Instrum Acq Met DA Meth	ename Type lent Namo hod hod	PS2 Una e Una Defa	547_01 vailable vailable ault.m	.d Sa Po Us IR Co	mple Name sition er Name M Calibration mment	Status	Unavailable Unavailable Unavailable Success Sample informati unavailable	ion is	1:	
User C	hromato	ograms							/	
Frag	gmentor Vo	oltage	90	Collision Ener	gy O I	onization Mode	Esi			
x10 ⁷	+ TIC So	an PS25	47_01	.d						
6	1				* 4.128				1	
5										
4										
2										
5										
2										
1-										
l										
	0.5	1 1.5	2	2.5 3 3.5 Co	4 4.5 5 unts vs. Acqu	5.5 6 isition Time (n	6.5 / /.5 nin)	8 8.5 9	9.5 10	
Integratio	on Peak Lis	st								
Peak	Start	RT	End	Height	Area	Area %]			
1	4,039	4,128	4,4	6206416	1 514376014	4 100)			
x10 ³ 2-	DAD1 - /	A:Sig=21	0,4 PS	2547_01.d	* 4.067					
1.5-										
1-										
0.5										
0.5										
0-		1								
Integratio) 0.5	1 1.5	2	2.5 3 3.5 Respon	4 4.5 5 se Units vs. A	5 5.5 6 Acquisition Tim	6.5 7 7.5 ne (min)	8 8.5 9	9.5 10	
Peak	Start	RT	End	Height	Area	Area %	1			
1	3.967	4.067	4.3	2239.0	4 11725.8	1 100	5			
User S	pectra						-			
Spectrum	Source				Fragmentor V	oltage	Collision Energy	Ionizat	tion Mode	-
Peak (1) in	+ TIC Sca	n"			90		0	2011/201	Esi	
	+ Scan (4 061-4 3	874 mi	n 43 scans) P	S2547 01 d 9	Subtract (1)				
x10 °	Court (202.20					1	



1



--- End Of Report ---





Qualitative Analys

Data Filename Sample Type **Instrument Name** Acq Method **DA Method** Default.m

PS2531_01.d Unavailable Unavailable

Sample Name Position **User Name IRM Calibration Status** Comment



User Chromatograms



Integratio	on Peak Lis	t								
Peak	Start	RT	End	Height	Area	Area %				
1	6,541	6,658	6,893	122152087	781421380	100				
x10 3	DAD1 - A	A:Sig=21	0,4 PS25	531_01.d						
1.2						* 6.60)7			
1-										
0.8										
0.6										
0.4										
0.2										
0	· · · · · · · · · · · · · · · · · · ·									
-0.2										
	0.5	1 1.5	2 2.5	3 3.5 4	4 4.5 5	5.5 6 6.5	7 7.5	8 8.5	9	9.5

3.5 4 4.5 5 5.5 6 6.5 7 7.5 8 8.5 9 Response Units vs. Acquisition Time (min) 3

Integration Peak List

Peak	Start	RT	End	Height	Area	Area %
1	6,527	6,607	6,707	1473,02	5461,03	100

User Spectra



10



--- End Of Report ---

1ª

"SK_zh10_04192023

bbo_1H_bar DMSO /v nmrsu 16

¹H NMR (500 MHz, DMSO) δ = 7.74 (d, *J*=8.2, 1H), 7.34 (d, *J*=8.2, 1H). 2.64 (q, *J*=7.6, 1H), 2.51 (dd, *J*=43.0, 17.7, 1H), 2.25 (dd, *J*=60.7, 16.0, 1H). 1.19 (t, *J*=7.6, 2H).



12g

0



Data Filename Sample Type		ZH-10_t1.d Sample	Sample Name Position	ZH-10 Vial 2	12g
Acq Method IRM Calibration State	ıs	All_2021_kol 1-2.m Not Applicable	Acquired Tim DA Method	e 5/12/2023 10:01:36 AM Defaul1t.m	o N N N N N N N N N N N N N N N N N N N
Sample Group Stream Name	LC 1	1	Info. Acquisition SW Version	6400 Series Triple Quadrupole 10.0 (127)	O NH2

User Chromatograms







--- End Of Report ---





bbo_13CF_bar DMSO /v nmrsu 3



Data Filename Sample Type		ZH-9_t1.d Sample	Sample Name Position	e ZH-9 Vial 2	12h
Instrument Name Acq Method IRM Calibration St Comment	atus	Instrument 1 All_2021_kol 1-2.m Not Applicable	User Name Acquired Tim DA Method	e 5/12/2023 9:41:38 AM Defaul1t.m	
Sample Group Stream Name	LC 1		Info. Acquisition SW Version	6400 Series Triple Ouadrupole 10.0 (127)	

User Chromatograms



Integration Peak List



			and the Marda
Spectrum Source	Fragmentor Voltage	Collision Energy	ESI
Peak (1) in "+ TIC Scan"	135	0	



Spectrum Source

Peak (1) in "DAD1 - B:Sig=220,4 Ref=off"



--- End Of Report ---

SK_zh-20_05172023

-7.72

Т



4.6 f1 (мд)





Data Filename Sample Type		ZH20_01.d Sample	Sample Name Position	2H20 Vial 2	13a 🧹
Instrument Name Acq Method IRM Calibration St	atus	Instrument 1 All_2021_kol 1-2.m Not Applicable	User Name Acquired Tim DA Method	e 5/29/2023 10:43:36 AM Defaul1t.m	o N
Comment Sample Group Stream Name	LC 1		Info. Acquisition SW Version	6400 Series Triple Ouadrupole 10.0 (127)	

User Chromatograms





User Spectra

			Tenination Mode
Spectrum Source	Fragmentor Voltage	Collision Energy	ESI
Peak (1) in "+ TIC Scan Sub"	135	0	



Spectrum Source

Peak (1) in "DAD1 - A:Sig=210,4 Ref=off Sub"



--- End Of Report ---







Data Filename		ZH19C_01.d Sample	Sample Name Position	ZH19C Vial 2	13b
Instrument Name Acq Method IRM Calibration St	atus	Instrument 1 All_2021_kol 1-2.m Not Applicable	User Name Acquired Time DA Method	5/29/2023 10:27:13 AM Defaul1t.m	o-J-
Comment					
Sample Group Stream Name	LC 1		Info. Acquisition SW	6400 Series Triple Quadrupple 10.0 (127)	ONH ₂

User Chromatograms





Spectrum Source Peak (1) in "+ TIC Scan Sub"



---- End Of Report ----

δ¹H NMR (500 MHz, DMSO) δ = 7.71 (d, *J*=7.8, 1H), 7.33 (d, *J*=7.5, 1H), 2.64 (dd, *J*=14.2, 6.9, 1H), 2.58 (d, *J*=18.0, 1H), 2.44 (d, *J*=17.7, 1H), 2.34 (d, *J*=16.1, 1H), 2.16 (d, *J*=16.0, 1H).



'SK_zh-16_05102023

bbo_13CF_bar DMSO /v nmrsu 10



Data Filename Sample Type		ZH6C_01.d Sample	Sample Nam Position	vial 2	13c
Instrument Name Acq Method IRM Calibration St Comment	tatus	Instrument 1 All_2021_kol 1-2.m Not Applicable	Acquired Tin DA Method	ne 5/29/2023 10:10:20 AM Defaul1t.m	
Sample Group Stream Name	LC 1	I A V	nfo. Acquisition SW Version	6400 Series Triple Quadrupole 10.0 (127)	

User Chromatograms



Integration Peak List

1



Start End Height Peak RT 8,079 7,805 7,892

User Spectra			
Spectrum Source	Fragmentor Voltage	Collision Energy	Ionization Mode
Peak (1) in "+ TIC Scan Sub"	135	0	ESI

4904,71

100

Area

1077,45



Spectrum Source

Peak (1) in "DAD1 - A:Sig=210,4 Ref=off Sub"



--- End Of Report ---

1







bbo_13CF_bar DMSO /v nmrsu 12



Data Filename Sample Type	ZH5C_01.d Sample	Sample Name Position User Name	ZH5C Vial 2	13d
Instrument Name Acq Method IRM Calibration Status Comment	All_2021_kol 1-2.m Not Applicable	Acquired Time DA Method	5/29/2023 9:54:12 AM Defaul1t.m	O N CO ₂ Et
Sample Group Stream Name LC 1	Info Acq Vers	u isition SW 64	00 Series Triple Iadrupole 10.0 (127)	

User Chromatograms





Peak	Start	RT	RT End	Height	Area	Area %	
1	7 759	7.846	8.032	1738,86	9135,32	100	

User Spectra

•				
Spectrum Source	Fragmentor Voltage	Collision Energy	Ionization Mode	
Peak (1) in "+ TIC Scan"	135	0	ESI	



Spectrum Source



--- End Of Report ---





Data Filename	HP-9_t1.d	Sample Name	HP-9	15 0 CI
Sample Type	Sample	Position	Vial 3	
Instrument Name	Instrument 1	User Name		N
Acq Method	All_2021_kol 1-6.m	Acquired Time	2/27/2024 12:49:38 PM	N_1 $NH_2 \downarrow O$
IRM Calibration Status	Not Applicable	DA Method	Defaul1t.m	
Comment				
Sample Group	In	Info.		$H_2N^{\prime} N^{\prime} O^{\prime} \sim$
Stream Name LC 1	Ac	quisition SW 64	100 Series Triple	
	Ve	rsion Qu	uadrupole 10.0 (127)	

User Chromatograms



2.5 3 3.5 4 4.5 5 Response Units vs. Acquisition Time (min)

3868,69

Area

Fragmentor Voltage

135

Area %

100

Collision Energy

0

Agilent Technologies

0

Peak

0.5

2,53

RT

Integration Peak List

User Spectra

Spectrum Source

Start

Peak (1) in "+ TIC Scan Sub"

1

2,73

1.5

End

2,96

2

Height

701,22

7

6.5

Ionization Mode

ESI

6



Spectrum Source

Peak (1) in "DAD1 - A:Sig=210,4 Ref=off Sub"



--- End Of Report ---






User Chromatograms





Peak List z Abund m/z 1 47228.23 255.1 1274338.25 400.1 1 300627.16 401.1 1 1 44883.64 402.1 1 197416.97 422.1 423.1 1 50234.55 19844.97 799.3 1 251334.08 1 821.3 822.3 1 125602.36 1 35859.36 823.3



---- End Of Report ----

Agilent Technologies







User Chromatograms



Integration Peak List

1 64		the second s	Contraction of the second s		
1 0,1	6,21	6,5	34068581,15	268022536.3	100



User Spectra

Spectrum Source Peak (1) in "+ TIC Scan Sub"	Fragmentor Voltage	Collision Energy	Ionization Mode		
	135	0	ESI		



--- End Of Report ---







User Chromatograms



Integration Peak List

Peak	1	Start	RT		End	Height		Area		Area	%				
	1	6,4		6,51	6,97	4248303	30,16	3	07213165,9	Ð	100				
x10 ³	[DAD1 - A	:Sic	1=210	.4 Ref=c	off HP-10 k01	.d Su	ubtract							
2	-	Λ													
1.5	+														
1	+										+				+
0.5	+						-			* 6 / 1					-
0															
	L	0.5	1	1.5	2 2.5	3 3.5 Response	4 4 Units	.5 5 vs. Acc	5.5 6 uisition Ti	6.5 me (mii	7 7.5 1)	8	8.5	9	9.5
ntegra 'eak	tio	on Peak Start	List RT		End	Height	Area	a Ar	ea %	1					
	÷	6.25		6 41	6 5	102.41	6	04.0	10	7					

User Spectra

Spectrum Source	Fragmentor Voltage	Collision Energy	Ionization Mode
Peak (1) in "+ TIC Scan Sub"	135	0	ESI



--- End Of Report ---

Agilent Technologies





Data Filename Sample Type Instrument Name Acq Method	e	HP-1_02.d Sample Instrument 1 All_2021_kol 1-2.m	Sample Nam Position User Name Acquired Tin	ne HP-1 Vial 2 ne 4/13/2023 10:20:40 AM	15e	0
Comment	Status	Not Applicable	DA Method	Defaul1t.m	٢	i j
Sample Group		Info			N NH2	
Sucan Name	LC 1	Acqu Vers	iisition SW ion	6400 Series Triple Quadrupole 10.0 (127)	H ₂ N N	0"

User Chromatograms





Spectrum Source

Peak (1) in "DAD1 - C:Sig=260,4 Ref=off"



--- End Of Report ----





Data Filename		HP-11_k01.d	Sample Nam	e HP-11	15f /
Sample Type		Sample	Position	Vial 2	
Instrument Name		Instrument 1	User Name		
Acg Method		All_2021_kol 1-6.m	Acquired Tim	e 2/19/2024 12:02:29 PM	0-
IRM Calibration Sta	tus	Not Applicable	DA Method	Defaul1t.m	N
Comment					N NH ₂ 0
Sample Group		en kan vianet en	Info.		
Stream Name	LC 1		Acquisition SW Version	6400 Series Triple Quadrupole 10.0 (127)	H ₂ N N O

User Chromatograms



Integration Peak List



Spectrum Source Peak (1) in "+ TIC Scan Sub" **Fragmentor Voltage** 135

ESI



--- End Of Report ---

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